ISOCAESPITOL, A NEW HALOGENATED SESQUITERPENE FROM LAURENCIA CAESPITOSA[†]

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Abstract—The halogenated sesquiterpene isocaespitol (1) has been isolated from the marine alga Laurencia caespitosa Lamx and its structure determined by X-ray crystallography.

In previous communications¹² we have established the structure of caespitol by relating it with isocaespitol (1), both compounds being terpenoids which contain bromine and chlorine, isolated from the marine alga *Laurencia caespitosa* Lamx (Rhodomelaceae). The present paper describes the chemical study of 1 and the confirmation of its structure by X-ray analysis.

RESULTS AND DISCUSSION

The air-dried, pulverised plant was extracted with ether, the ether concentrate was washed successively with dil KOH, dil HCl and water, and the neutral oil thus obtained was carefully chromatographed on standard silica gel. The ether-benzene (9:1) eluent contained

*Part XII in the series "Marine Natural Products of the Atlantic Zone". For Part XI see A. G. González, J. Darias, J. D. Martín and M. Norte, Tetrahedron Letters 3951 (1974). isocaespitol (1); 0.06% dry weight alga) as a colourless oil which slowly crystallized, m.p. $92-93^{\circ}/n$ -hexane, $[\alpha]_{D} = -15^{\circ}$.

Isocaespitol (1), was analysed for $C_{15}H_{25}O_2Br_2Cl$ by mass spectrometry [*m/e* 434, 432, 430 (M⁺); high resolution *m/e* 353.073 ($C_{15}H_{25}O_2^{30}Br^{35}Cl$ requires: 353.070), 237.187 ($C_{15}H_{25}O_2$ requires 237.187)], $\nu_{max}(KBr)$ 3500, 3320, 1650, 950, 830, 810 and 720 cm⁻¹. The NMR spectrum (100 MHz, CDCl₃, τ -scale) and the spin decoupling study indicate the presence of two one-proton signals at 5.54 (s, *W*1/2 5 Hz) and 5.70 (4 lines, X of ABX) corresponding to protons α to halogens, 6.53 (t, *J* 3 Hz, shifted to 5.36 on acetylation) of one proton α to an OH group, and an AB part of an ABXY pattern centred at 7.63 and 7.72. In the upfield region appear signals for four tertiary Me groups at 8.08, 8.62, 8.69 and 8.84. Acetylation of 1 gave the monoacetate 2, from which isocaespitol was regenerated by mild saponification. The mass spectra of

	x ¹	Y	2	8150 ²	B22	B ₃₃	B12	^B 13	^B 2 3
BR1	.8623 (4)	.595(1)	. 5265 (4)	3.2(2)	9.7(5)	4.1(3)	-0.9(6)	0.2(2)	-2.2(4
BR2	.1107 (4)	0	.0806(5)	2.7(2)	8.3(5)	10.0(4)	-4.9(6)	0.7(2)	-1.8(4
CL	.843 (1)	.259(3)	.203(1)	4.5(7)	8.5(14)	6.3(8)	-2.5(14)	2.1(6)	-3.9(9
0,	. 398 (2)	.374 (4)	.183(2)	3.1(7)					
0,	. 532 (2)	.031(4)	. 167 (2)	2.8(6)					
н ₂ 0	.481(2)	.682(4)	.051(2)	4.2(7)					
ci	.708(3)	.238(6)	.358(3)	2.3(10)					
C2	.826(4)	.321(8)	.346(4)	5.0(13)					
C3	.842 (3)	. 563 (7)	. 352 (3)	1.5(8)					
C4	.728(3)	.649(7)	.289(3)	2.3(9)					
C5	.614 (3)	. 583 (9)	.311(3)	3.6(10)					
C6	.602(3)	.380(7)	.281(3)	2.3(10)					
C7	.477(3)	.268(7)	.280(3)	2.4(10)					
C8	.476(3)	.061(7)	. 262 (3)	2.9(10)					
C9	.344 (3)	011(7)	. 229 (3)	2.7(10)					
C10	.274 (3)	.121(8)	.123(3)	3.5(10)					
c11	.271(4)	.311(9)	.135(4)	4.7(13)					
C12	.197(3)	.408(6)	.223(3)	2.7(10)					
C13	.233(3)	.426(6)	.019(3)	2.7(10)					
C14	.444 (3)	.325(7)	. 398 (3)	2.0(9)					
C15	.958(3)	. 624 (8)	.331(3)	4.1(11)					

Final Atom Coordinates for Isocaespitol

1) Standard deviations of last significant figures in parenthesis.

2) B in \Re^2 or B_{ij} converted to the same units as B using the equation B_{ij} = 4 b_{1j}/a₁^{*}a₁^{*}.

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the alcohol and acetate showed base peaks at (257, 255, 253) and (299, 297, 295) respectively, arising from the fragment [a] as is shown in 1. The C-10 halogen was located when it was shown that treatment of the alcohol (1) with Jones reagent gave the unstable monoketone (3) which easily loses HBr to yield the α,β -unsaturated ketone (4). On the basis of the aforesaid, the relative position of the chlorine and bromine at C-3 and C-4 could not be assigned.

In order to definitively elucidate the structure, a single-crystal of isocaespitol was subjected to X-ray diffraction analysis. One suitable crystal was obtained by recrystallization from carbon tetrachloride. The compound crystallized in the monoclinic system, with systematic extinctions in its diffraction pattern for OkO with k = 2n + 1. Since the sample of isocaespitol used is optically active, the space group is uniquely fixed as P2₁. Unit cell constants were determined from a least squares fit of 12 carefully centred reflections measured on a Picker FACS-II automated differactometer (MoK_{a1}; $\lambda = 0.70926$ Å) to be: a = 11.548 (13), b = 6.961 (9), c = 11.859 (12) Å and $\beta = 104.27$ (3)°.

Three-dimensional intensity data were collected on the above diffractometer, using MoK_a radiation made monochromatic by Bragg reflection from a graphite crystal, scanning reflections in the $2\theta - \theta$ mode at a scan rate of 1°/min and a scan range of 2.0°, and taking background counts of 10 sec at each end of the scan. Of 649 unique reflections having 2θ values less than 35°, 583 remained after rejection of those with intensities less than 1.5σ . These were put on a common scale using the intensities of three standards (collected every 100 reflections) and corrected for Lorentz-polarization effects. The structure was solved by the heavy atom method, successive cycles of Fourier synthesis, and full-matrix least squares refinement. Ambiguities remained concerned the location of OH and O atoms as well as the identity of a foreign peak in a difference synthesis made very late in the refinement. Careful analysis of the temperature factors associated with each atom led to satisfactory locations for all oxygens in the natural product and a very strong indication that the foreign peak represents a water of crystallization. The H-bonding scheme between the alleged water and the OH and ether moieties reinforces that conclusion. Even though anomalous scattering from the Br and Cl atoms was carefully considered we were unable to arrive at an absolute configuration.

In 1 we show the geometry of isocaespitol. The final weighted residue, wR, was 0.087, and the standard deviation of an observation of unit weight is 1.72. All

bond distances and angles agree well with generally accepted values within experimental error.³

EXPERIMENTAL

The m.ps were determined on a Kofler block and are uncorrected. UV spectra, recorded with a Beckman DB-GT spectrophotometer, refer to EtOH solns. IR spectra, measured with a Unicam SP1100 spectrometer in KBr if not otherwise indicated. NMR spectra were taken with a Perkin-Elmer R-12 (60 MHz) or a Varian HA-100 (100 MHz) instrument for CDCl₃ solns with TMS as internal standard, W1/2 refers to the width of a band at half height. MS were recorded on an A.E.I. MS-902 or a Hitachi-Perkin-Elmer RMU-7 instrument. Column and dry column chromatography was performed on silica gel 0.2–0.5 and 0.063–0.20 mm respectively, TLC and PLC on silica gel G, the spray reagent was $6N H_2SO_4$. Petrol, refers to the fraction b.p. 40–60°. Anhys Na₂SO₄ was used for drying solns.

Isolation of isocaespital (1). Air dried seaweed (2.0 kg) was extracted with ether and the ether soln was concentrated in vacuo. The residue was washed successively with 5% KOH aq and then with 1N HCL to remove acidic and basic components. The neutral oil thus obtained (25 gr) was carefully chromatographed on standard silica gel. Fractions eluted with ether-benzene (9:1) on removal the solvents, left a crystalline substance. Recrystallization from *n*-hexane gave 1 (1.2 g) as monoclinic crystals, m.p. 92-93°, $[\alpha] - 15$ (c, 0.12). (Found: C, 40.04; H, 60.2 C₁₃H₂₃O₂Br₂Cl. H₂O requires: C, 39.94; H, 5.99%). IR: see text. NMR: see text. MS: *m/e* (%) 434, 432, 430 (M⁻, 2): 355, 353, 351 (16): 268, 266 (70): 257, 255, 253 (100): 237 (5): 223, 221 (94): 205, 203 (22): 175, 173 (93): and 137, 135, 133 (60).

Acetylation of isocaespitol. Isocaespitol (30 mg) was allowed to react overnight in a soln of Ac₂O (2 ml) in pyridine (1 ml). The soln was poured into water and the product isolated with ether, purified by elution from silica gel with benzene, and crystallised from *n*-hexane as long needles, m.p. 125–128^c. (Found: C, 41·50; H, 5·94. C₁₇H₂₇O₃Br₂Cl. H₂O requires: C, 41·42; H, 5·88%); IR: 3500, 2980, 2935, 1735, 1380, 820, 790 and 720 cm⁻¹; NMR: 8·77, 8·66, 8·60, 8·08, 7·90 (each 3H, s), 5·89 (1H, q, *J* = 12 and 6 Hz), 5·54 (1H, bs, *W*1/2 = 6 Hz) and 5·36 (1H, t, *J* = 3 Hz). MS: *mle* (%) 476, 474, 472 (M^{*}, 3); 397, 395, 393 (20); 310, 308 (64); 299, 297, 295 (100); 279 (16); 265, 263 (80); and 175, 173 (92).

Oxidation of isocaespitol. To a stirred soln of 1 (100 mg) in Me₂CO (20 ml) at room temp. Jones reagent (0.3 ml) was added dropwise till a permanent orange colour resulted. After a further 5 min stirring the mixture was worked up giving the unstable ketone 3 (94 mg); IR: 2985, 2930, 1710, 900 and 710 cm⁻¹; NMR: 8-72, 8-58, 8-54 and 8-08 (each 3H, s), 7-04 (1H, d, J = 8 Hz), 7-03 (1 H, d, J = 10 Hz), 5-76 (1H, q, J = 10 and 8 Hz) and 5-54 (1H, bs, W1/2 = 4 Hz). Without further purification, 3 was chromatographed on alumina yielding 4 (54 mg) as a crystalline substance. Recrystallization from n-hexane gave plates m.p. 67–68°: UV 224 nm (ϵ 16,900); IR: ν_{max} (CCL) 2980, 2930, 1685, 1375, 1100, 1070, 1020, 820 and 730 cm⁻¹; NMR: 8-70 (3H, s), 8-60 (eH, s), 8-09 (3H, s), 5-43 (1H, bs, W1/2 = 6 Hz), 4-19 and 3-26 (each 1H, d, J = 11 Hz). MS: m/e (%) 353, 351, 349 (M⁻, 2); 257, 255, 253 (100); 219, 217 (20); 175, 173 (48) and 139 (34).



REPERENCES

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